

Association of Malaria with Thrombocytopenia

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In this study, 250 blood samples submitted for malaria investigation were studied microscopically for malarial parasites and platelet count. All samples were additionally analyzed for platelet count with automated haematology analyzer. Thirty seven (37) samples were found to be malaria positive microscopically. Out of 37 cases with malaria positive microscopically, thrombocytopenia was observed in 24 (64%) cases of malaria. So there is association of thrombocytopenia with malaria.

Key words: Malaria, thrombocytopenia, *P. falciparum*

300-500 million people are infected by malaria yearly¹ with the mortality rate of 1%². It is epidemic and endemic in Africa, Central and South America, the Middle East and parts of Asia which have hot, humid environment which is ideal for breeding of anopheles mosquito and is transmitted by the bite of infected female anopheles mosquito. It is also transmitted by blood transfusion, transplacentally and between drug addicts by reusing syringes³.

Four species of plasmodia i.e. *P. vivax*, *P. ovale*, *P. falciparum* and *P. malariae* can cause malaria which is distinguishable on peripheral blood smear⁴. Plasmodium sporozoites are injected by the bite of female mosquito, reach liver, multiply there and released after 1-2 weeks to infect red cells. Severe cases of malaria are seen in *falciparum* infection. The severity of malaria may be determined by the magnitude of parasitaemia. The malaria is usually presented with febrile paroxysms, malaise and anemia². The main hematological findings in patient's blood are anemia, thrombocytopenia, variable (low, normal, high) white cell (WBC) count, bleeding and parasitaemia^{2,5}. Malaria is still common in oasis and coastal areas of the Saudi Arabia. Expatriate work force also imports malaria from their home countries especially endemic areas of malaria⁶.

Materials and methods:

Two hundred and fifty (250) adult subjects suspected of malaria were selected from Riyadh Medical Complex. Thirty seven (37) were found positive for malarial infection microscopically by thick and thin smears. Platelet counts were performed manually⁷ as well as with automated hematology analyzer (Cell Dyn 3700). All slides were stained by Giemsa method⁸. Identification and level of parasitemia was done for each positive case⁹.

Results:

In this study two types of plasmodium species, *P. falciparum* and *P. vivax* were found on thick and thin smear. There were 34(92%) cases of *falciparum* and 3(8%) cases of *vivax*. The comparison of positive and negative cases is given in Table 1

Five (20.1%) patients had platelet count less than $50 \times 10^9/L$, eleven (45.1%) had thrombocytopenia in the range of $50-100 \times 10^9/L$ and eight (34.8%) had thrombocytopenia in the range of $100 - 150 \times 10^9/L$. Out of these five patients, three have parasitemia in the range of 3-10%. Maximum parasitemia was found to be 10% while 0.1% was the lowest. There were 8 (21.6%) patients who have parasitemia of 1% or above. These patients have platelet count of $60 \times 10^9/L$ or less and this high parasitemia is inversely related to platelet count. A low platelet count was associated with high parasitaemia ($p < 0.05$).

Table 1: Comparison of platelet count in malaria positive and negative cases with microscopy and hematology analyzer

Methods	Platelet count in malaria positive cases	Platelet count in malaria negative cases
Microscopy	117 ± 35.06	285.6 ± 41.01
Hematology analyzer	137 ± 48.13	292.3 ± 40.3

Discussion:

Thrombocytopenia is a well documented finding in *falciparum* malaria and in mixed *falciparum/vivax* infection^{11,12}. A platelet count less than $150 \times 10^9/L$ was considered thrombocytopenia but is not associated with adverse outcome¹³. Thrombocytopenia is considered as an important indicator of malaria¹⁴. Maximum thrombocytopenia occurs on the fifth or sixth day of infection and gradually returns to normal within 5-7 days after parasitemia has ceased¹⁵. In the present study thrombocytopenia of less than $150 \times 10^9/L$ was found in 24 (65%) of the malaria cases. Mean platelet count in *P. falciparum* infection was $141 \times 10^9/L$ in hematology analyzer while on microscopy the mean platelet count was $120 \times 10^9/L$.

Thrombocytopenia has been observed in 60-80% of both *P. falciparum* and *vivax* infection¹⁵. The shortened life span of platelet is 2-3 days in comparison to 7-10 days in normal controls^{15,16}. The mechanism of thrombocytopenia in malaria is still unclear. Fajardo and Tallent¹⁷ suggested a direct lytic effect of parasite on platelets. Both non-immunological destruction¹⁸ and immunological mechanism involving platelet specific antibodies¹⁹ have

been demonstrated. Mohanty et al²⁰ suggested that thrombocytopenia in malaria is partly immune mediated. During malarial infection, initial hyperactivity results in aggregation and later hypoactivity of platelets causes intravascular lysis. There is peripheral destruction and consumption of platelet in infected persons. Srichaikul¹⁸ noted that despite thrombocytopenia, the number of megakaryocytes in the bone marrow remained adequate or increased in malarial infection.

Scott et al¹⁴ found that two third patients with parasitemia above 10% had platelet count $< 50 \times 10^9/L$. Ladhani et al¹³ found that a low platelet count is associated with parasite density but not with bleeding problem or mortality. Geradin et al²¹ found an association between thrombocytopenia and either severity or prognosis in childhood falciparum. Low parasitemia may lead to the misdiagnosis of malaria and may delay the treatment²². Thus cases of pyrexia of unknown origin and thrombocytopenia should be investigated thoroughly for malaria.

Thus screening complete blood count can be a rapid and inexpensive yet valuable component in the diagnostic investigation of any patient, particularly the patient with pyrexia of unknown origin and thrombocytopenia.

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